

# Finished Genome Sequence of *Collimonas arenae* Cal35

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**We announce the finished genome sequence of soil forest isolate *Collimonas arenae* Cal35, which comprises a 5.6-Mbp chromosome and 41-kb plasmid. The Cal35 genome is the second one published for the bacterial genus *Collimonas* and represents the first opportunity for high-resolution comparison of genome content and synteny among collimonads.**

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Collimonads are oligotrophic, chitinolytic, rhizosphere-competent soil bacteria with antifungal, mycophagous, and mineral-weathering properties (1). To date, three species of *Collimonas* have been described (2, 3), but the only *Collimonas* genome available is that of the Dutch dune soil isolate *C. fungivorans* Ter331 (*Cf*Ter331, GenBank accession no. CP002745 for the chromosome and EU315244 for plasmid pTer331). The *Cf*Ter331 genome has been instrumental in describing the *Collimonas* chitinolytic system (4), the genes underlying the production of antifungal polyene-like compounds called collimomycins (5), the molecular interactions with model fungus *Aspergillus niger* (6), and the phenotypic variation among other collimonads isolated from the same dune soil as Ter331 (7). However, with only one *Collimonas* genome sequence available to date, intrgeneric comparison at single-nucleotide resolution has not yet been feasible. Here, we report the finished genome sequence of *Collimonas arenae* Cal35 (*Ca*Cal35), which was recovered from a forest soil in California (8).

Genomic DNA of *Ca*Cal35 was extracted from an overnight culture using a DNeasy Blood and Tissue kit (QIAGEN, Valencia, CA). A 10-kb PacBio RS II-compatible library was constructed by the UC Davis Genome Center and sequenced on 3 single-molecule real-time (SMRT) cells using P4-C2 chemistry. *De novo* assembly was performed with the help of SMRT Analysis software v2.2.0 (Pacific Biosciences) featuring HGAP 2 (9), and subsequent correction with quiver in addition to Gepard v1.30 (10) to reveal two circular replicons: a 5,603,532-bp chromosome (G+C content 56.15%; 85.54× coverage) and a 41,440-bp plasmid (G+C content 50.54%; 73.21× coverage) which we designated pCal35. Gene prediction by RAST (11) revealed 5,019 coding sequences, 54 tRNA genes, and 3 rRNA operons (5S, 16S, 23S) on the chromosome, in addition to 57 coding sequences on pCal35.

The chromosome of *Ca*Cal35 appears mostly syntenic with that of *Cf*Ter331, with the exception of one 2.75-Mbp inversion centered on the origin of replication. *Ca*Cal35 shares 69% of its genes with Ter331, of which 80% are >80% identical at the predicted amino acid level. The chitinolytic system of *Ca*Cal35 is very similar to that of *Cf*Ter331, but *Ca*Cal35 possesses at least two

additional chitinase genes. Clearly missing from the *Ca*Cal35 genome is gene cluster K which is responsible for the production of collimomycins (5–7). Interestingly, in *Cf*Ter331, this gene cluster is located near one of the chromosome inversion points. Unrelated to pTer331 (12), plasmid pCal35 carries several genes with high coding similarity to plasmids from plant-pathogenic species of *Xanthomonas* (for example, pXAC64, pXCV38, pXcB) and *Ralstonia* (pRSC35). In addition, it harbors a stretch of DNA with high similarity to plasmid 2 from *Polaromonas* JS666 and coding for tRNA 2-selenouridine synthase, alkane 1-monooxygenase, and a LuxR-type regulator.

**Nucleotide sequence accession numbers.** The Cal35 chromosome and plasmid sequences are available under GenBank accession numbers [CP009962](https://www.ncbi.nlm.nih.gov/nuclink/CP009962) and [CP009963](https://www.ncbi.nlm.nih.gov/nuclink/CP009963), respectively.

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Assembly and annotation of the *Ca*Cal35 genome sequence were performed by V.C.L.d.J. and J.J.W. (in the lab of J.H.J.L., visiting from the group of W.-L.D. as part of the ongoing UCD-NCHU collaboration). Interpretation of the annotation data was done by J.H.J.L., who also drafted the manuscript with inputs, edits, and final approval by all other authors. This is NIOO-KNAW publication number 5741.

## REFERENCES

1. Leveau JHJ, Uroz S, de Boer W. 2010. The bacterial genus *Collimonas*: mycophagy, weathering and other adaptive solutions to life in oligotrophic soil environments. *Environ Microbiol* 12:281–292. <http://dx.doi.org/10.1111/j.1462-2920.2009.02010.x>.
2. de Boer W, Leveau JHJ, Kowalchuk GA, Klein Gunnewiek PJA, Abeln ECA, Figge MJ, Sjollem K, Janse JD, van Veen JA. 2004. *Collimonas fungivorans* gen. nov., sp. nov., a chitinolytic soil bacterium with the ability to grow on living fungal hyphae. *Int J Syst Evol Microbiol* 54:857–864. <http://dx.doi.org/10.1099/ijs.0.02920-0>.
3. Höppener-Ogawa S, de Boer W, Leveau JHJ, van Veen JA, de Brandt E,

- Vanlaere E, Sutton H, Dare DJ, Vandamme P. 2008. *Collimonas arenae* sp. nov. and *Collimonas pratensis* sp. nov., isolated from (semi-)natural grassland soils. *Int J Syst Evol Microbiol* 58:414–419. <http://dx.doi.org/10.1099/ijss.0.65375-0>.
4. Fritsche K, de Boer W, Gerards S, van den Berg M, van Veen JA, Leveau JHJ. 2008. Identification and characterization of genes underlying chitinolysis in *Collimonas fungivorans* Ter331. *FEMS Microbiol Ecol* 66: 123–135. <http://dx.doi.org/10.1111/j.1574-6941.2008.00547.x>.
  5. Fritsche K, van den Berg M, de Boer W, van Beek TA, Raaijmakers JM, van Veen JA, Leveau JHJ. 2014. Biosynthetic genes and activity spectrum of antifungal polyynes from *Collimonas fungivorans* Ter331. *Environ Microbiol* 16:1334–1345. <http://dx.doi.org/10.1111/1462-2920.12440>.
  6. Mela F, Fritsche K, de Boer W, van Veen JA, de Graaff LH, van den Berg M, Leveau JHJ. 2011. Dual transcriptional profiling of a bacterial/fungal confrontation: *Collimonas fungivorans* versus *Aspergillus niger*. *ISME J* 5:1494–1504. <http://dx.doi.org/10.1038/ismej.2011.29>.
  7. Mela F, Fritsche K, de Boer W, van den Berg M, van Veen JA, Maharaj NN, Leveau JHJ. 2012. Comparative genomics of bacteria from the genus *Collimonas*: linking (dis)similarities in gene content to phenotypic variation and conservation. *Environ Microbiol Rep* 4:424–432. <http://dx.doi.org/10.1111/j.1758-2229.2012.00336.x>.
  8. Uroz S, Tech JJ, Sawaya NA, Frey-Klett P, Leveau JHJ. 2014. Structure and function of bacterial communities in ageing soils: insights from the Mendocino ecological staircase. *Soil Biol Biochem* 69:265–274. <http://dx.doi.org/10.1016/j.soilbio.2013.11.002>.
  9. Chin CS, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. *Nat Methods* 10:563–569. <http://dx.doi.org/10.1038/nmeth.2474>.
  10. Krumsiek J, Arnold R, Rattei T. 2007. Gepard: a rapid and sensitive tool for creating dotplots on genome scale. *Bioinformatics* 23:1026–1028. <http://dx.doi.org/10.1093/bioinformatics/btm039>.
  11. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <http://dx.doi.org/10.1186/1471-2164-9-75>.
  12. Mela F, Fritsche K, Boersma H, van Elsas JD, Bartels D, Meyer F, de Boer W, van Veen JA, Leveau JHJ. 2008. Comparative genomics of the pIPO<sub>2</sub>/pSB102 family of environmental plasmids: sequence, evolution, and ecology of pTer331 isolated from *Collimonas fungivorans* Ter331. *FEMS Microbiol Ecol* 66:45–62. <http://dx.doi.org/10.1111/j.1574-6941.2008.00472.x>.